

# Protective Role of Recombinant Human Erythropoietin in Kidney and Lung Injury Following Renal Bilateral Ischemia-Reperfusion in Rat Model

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## ABSTRACT

**Background:** Acute kidney injury (AKI) has been recognized as one of the most complex clinical complications in modern medicine, and ischemia/reperfusion (I/R) injury is well-known as a main reason of AKI. In addition, AKI leads to important systemic consequences such as acute lung injury. This study was designed to investigate the role of erythropoietin (EPO) on kidney function makers and tissue damage; and lung endothelial permeability and lung water content (LWC) in bilateral renal I/R injury model in rats.

**Methods:** Male Wistar rats were randomly divided into three groups of sham, I/R, and I/R treated with EPO (I/R + EPO) groups. The I/R and I/R + EPO groups were subjected to bilateral renal I/R injury; however, only the I/R + EPO group received EPO (500 IU/kg, i.p.) 2 h before ischemia surgery, and the same dose was continued once a day for 3 days after ischemia. The sham group underwent a surgical procedure without ischemia process.

**Results:** The blood urea nitrogen (BUN) and serum creatinine (Cr) levels, kidney tissue damage score (KTDS), and kidney weight (KW) per 100 g body weight significantly increased in I/R group (P < 0.05). EPO administration decreased levels of BUN and Cr significantly (P < 0.05), and KTDS and KW insignificantly (P = 0.1). No significant differences in kidney and serum levels of malondialdehyde, and lung vascular permeability and LWC were observed between the groups. The serum and kidney levels of nitrite were not significantly different between I/R and sham groups; however, administration of EPO increased the renal level of nitrite (P < 0.05).

**Conclusions:** EPO protected the kidney against I/R injury; however, it may not protect the lung tissue from the damage induced by renal I/R injury in rats.

Keywords: Erythropoietin, lung endothelial permeability, lung water content, rat

## INTRODUCTION

Acute kidney injury (AKI) is a common clinical syndrome that is induced by kidney ischemia.<sup>[1-5]</sup> Renal ischemia and

reperfusion (I/R) injury also is one of the common complications in clinical surgeries such as renal transplantation.<sup>[6]</sup> The I/R injury triggers an immune response and leads to both local and systemic inflammations. It disturbs renal function and immune system homeostasis.<sup>[7,8]</sup> The crosstalk between the renal injury and distant organs such as lung is one of the complicated processes having very complex mechanism.<sup>[9]</sup> The most important cause of the high rate mortality induced by AKI is related to the functional role of the pulmonary system.<sup>[10]</sup> The acute lung injury after AKI or I/R injury is featured by pulmonary vascular congestion, interstitial edema, focal alveolar hemorrhage, and inflammatory cell infiltration.<sup>[11]</sup> It is also reported that pulmonary vascular permeability increases after AKI or I/R injury.<sup>[12]</sup> Erythropoietin (EPO) is a 30.4 kD glycoprotein of class I cytokine consisting of 165 amino acids.<sup>[13,14]</sup> Potentially, it exhibits a powerful tissue-protective effect against I/R.<sup>[15]</sup> EPO has been the subject of many in vivo and in vitro researches as a nephroprotectant agent after kidney injury.<sup>[6,14,16-32]</sup> Although most data available support positive effects for EPO administration, some available evidence has been shown unfavorable effects of EPO in kidney injury.<sup>[18,21,31]</sup> However, less information have supported the pulmono-protective effects of EPO after I/R injury. Some published data demonstrated that EPO pretreatment in IRI rat model may attenuate renal and lung injuries,<sup>[17,33]</sup> but it seems that more pathological information is still needed. In this study, we attempted to examine the effect of EPO administration on kidney and lung tissues simultaneously by gathering biochemical and pathological data as well as lung vascular permeability (LP) in bilateral IRI rat model.

# **METHODS**

# Animals

A total of 19 adult male (weighting  $189 \pm 3.36$  g) Wistar rats (Animal Center, Isfahan University of Medical Sciences, Isfahan, Iran) were used in this study. Animals were housed under standard conditions with 12 h light/12 h dark cycle and had free access to water and food. Prior to experiment, the protocols were confirmed to be in accordance with the Guidelines of Animal Ethics Committee of Isfahan University of Medical Sciences.

# Drugs

EPO (recombinant human erythropoietin alpha) and Evans Blue were purchased from Pooyesh Darou Pharmaceutical Co. (Tehran, Iran) and Sigma (St. Louis, Missouri, USA), respectively.

## **Experimental protocol**

The animals were randomly divided into three experiment groups; sham, I/R, and I/R treated with EPO (I/R + EPO). The I/R + EPO group received EPO (500 IU/kg, i.p.) 2 h before the ischemia surgery, and administration of the drug continued for 3 days after ischemia. The I/R group followed the same regimen. Only it received saline instead of EPO. The sham group underwent the surgical procedure without ischemia process. To induce the IRI model, the animals in groups I/R and I/R + EPO were anesthetized by ketamine (75 mg/kg, i.p.) and xylaxine (10 mg/kg, i.p.). Two small incisions were made on the skin of the back of the animal, and the fascia was genteelly removed to appear the kidneys. The both kidney arteries and veins were clamped for 45 min. After removing the clamp, kidney reperfusion was performed. The animals were recovered after surgery for the following steps of an experiment. The animals in groups I/R and I/R + EPO respectively received daily saline and EPO 24, 48, and 72 h after renal IRI. The animal body weight was recorded on a daily basis. On day 3 (72 h post-IRI) and 2 h after the last injection, the animals were operated for the next step. After anaesthetization of the animals again, the trachea was cannulated by ventilation tube and the catheters were implanted into the carotid artery to obtain a blood sample and into the jugular vein for injection of Evans Blue (EB) solution (10 mg/kg). The right kidney was removed. To maintain stable anesthesia condition, animals received oxygen ventilation if needed. Finally, the rats were sacrificed 1 h after EB injection by potassium chloride solution. Tissue samples of lung and left kidney were obtained and fixed in 10% formalin solution for pathological assessment. Tissue samples of the lung were also collected to be investigated for edema and endothelial permeability. The removed right kidney was homogenated and centrifuged at 15,000g for 2 min, and the supernatant was used for biochemical measurements.

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## Measurements

Serum levels of creatinine (Cr) and blood urea nitrogen (BUN) were measured using quantitative kits (Pars Azmoon, Iran). Serum and kidney levels of nitrite (stable metabolite of nitric oxide [NO]) were measured using an ELISA assay kit (Promega Corporation, USA). Assessment of malondialdehyde (MDA) level in the serum and kidney was performed by the manual method. Briefly, a mixture of 500  $\mu$ l of the sample and 1000  $\mu$ l of 10% trichloroacetic acid (TCA) was centrifuged at 2000g for 10 min; then 500 µl of the supernatant was pulsed with 500 µl of 0.67% thiobarbituric acid (TBA). After 10 min of incubation in boiling water and then cooling, the absorbance was measured at 532 nm. Concentrations of MDA for serum and kidney samples were reported in µmol/1 and nmol/g tissue, respectively.

## Measurement of pulmonary water content

The lung tissue was kept in oven under 100°C until constant weight was obtained. The percentage of lung water content (LWC) was calculated as (wet lung weight – dry lung weight)\*100/wet lung weight.

### Measurement of LP

LP was measured by EB method that is described elsewhere.<sup>[34,35]</sup> Lung tissue was put into 4 cc formamide and kept in oven under 80°C for 24 h. The tissue EB was extracted by formamide, and then its absorbance was read at 623 nm to determine the tissue endothelial permeability ( $\mu$ g/g tissue) using standard curves.

### Histopathological procedures

The removed kidney and lung were fixed in 10% formalin solution, and then embedded in paraffin for histopathological staining. The hematoxylin and eosin stain was applied to examine the tissue injury. To consider the kidney damage, presence of tubular atrophy, hyaline cast, ischemic necrosis, vacuolization, and debris was evaluated. Cases with ischemic necrosis higher than 5% were excluded from the study. Based on the damage intensity, we scored the samples as 1-4 while score zero was assigned to normal tissue. To consider the lung tissue damage, presence of congestion, inflammation, and fibrosis were evaluated. Based on the damage intensity, the samples were scored in the range of 1-4 while score zero was assigned to normal tissue.

### Statistical analysis

The data are presented as mean  $\pm$  standard error of the mean. To compare the weight change, serum levels of BUN, Cr, MDA, and NO; kidney levels of MDA and NO, kidney weight (KW), and pulmonary permeability and edema were compared between the groups by the one-way analysis of variance, followed by least significant difference. Since, the scoring is qualitative; the Mann-Whitney or Kruskal-Wallis tests were applied to compare the pathology damage scores between the groups. *P* < 0.05 were considered statistically significant.

## **RESULTS**

# Effect of I/R injury on serum Cr and BUN levels

The BUN and Cr levels increased in the I/R group in comparison with the sham group (P < 0.05). In the presence of EPO, the serum concentrations of Cr and BUN were lower than I/R group (P < 0.05) [Figure 1].

# IRI effects on kidney tissue damage score and total KW

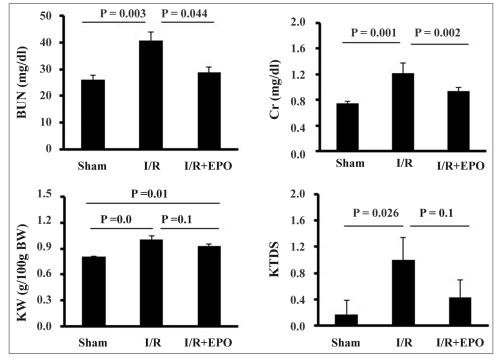
The KTDS and KW in gram per 100 g body weight in the I/R group significantly increased when compared to the sham group (P < 0.05). However, EPO administration reduced KTDS and KW in comparison with the I/R group (P = 0.1) [Figure 1]. The image of kidney tissue damage is demonstrated in Figure 2.

# Effect of I/R injury on serum and kidney tissue levels of MDA and nitrite, and bodyweight

The data for the serum and kidney tissue levels of MDA and nitrite, and bodyweight change is tabulated in Table 1. No significant differences were observed in the MDA and nitrite levels between the I/R and sham groups. However, EPO did not change the MDA level, but the kidney nitrite level in the I/R + EPO group was greater than that in the I/R group (P < 0.05). No bodyweight change was detected between the groups.

# Effect of I/R injury on LP, LWC, and lung tissue damage score

No significant difference in the LP and LWC were observed between the groups. However, EPO administration significantly increased the



**Figure 1:** Serum creatinine and blood urea nitrogen levels, total kidney weight per 100 g body weight, and kidney tissue damage score in the sham, ischemia/reperfusion (I/R), and I/R treated with erythropoietin groups

Table 1: Serum and kidney	y tissue levels of MDA and nitrite,	and body weight change	ge in the sham, I/R, a	and I/R+EPO groups

				_	
Group	Serum MDA	Kidney MDA	Serum nitrite	Kidney nitrite	Body weight change
	(µmol/l)	(nmol/g tissue)	(µmol/l)	(nmol/g tissue)	<b>(g)</b>
Sham	2.69±0.49	1.72±0.31	12.71±2.60	0.17±0.02	7.5±2.4
I/R	3.41±0.67	2.92±1.06	7.65±0.94	$0.14{\pm}0.01$	5.6±2.7
I/R+EPO	3.32±0.81	2.34±1.03	9.12±2.34	0.19±0.03*	6.5±2.4

\*Significant difference from the I/R group, P<0.05. MDA=Malondialdehyde, I/R=Ischemia/reperfusion, EPO=Erythropoietin

lung tissue damage when compared to the sham group (P < 0.05) [Figure 3]. The sample lung tissue damage is demonstrated in Figure 2.

### DISCUSSION

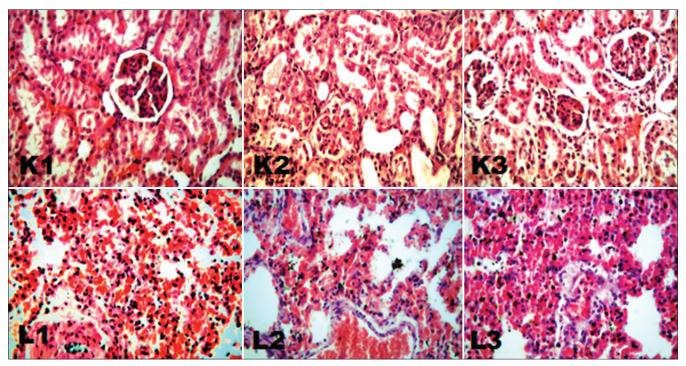
Findings of this study indicated that EPO has protective effects on renal injury induced by bilateral renal IR, without positive effects on lung injury. Our results are confirmed by the previous studies.<sup>[27,36-38]</sup> The increased KW in the IR group may be related to renal edema or cell proliferation in kidney tissue, which is in agreement with the reports of Forbes *et al.*<sup>[39]</sup> The protective effect of EPO against renal IRI may be associated with its antioxidant, anti-apoptotic, anti-inflammatory, and angiogenic properties.<sup>[13,15,36,37,40-42]</sup> The body weight loss induced by IR occurs due to inability of the kidney for salt and

water retention<sup>[43]</sup> or cachexia and polyuria.<sup>[12]</sup> It was also reported that EPO (500 U/kg) administration after renal IRI had no effect on bodyweight.<sup>[44]</sup>

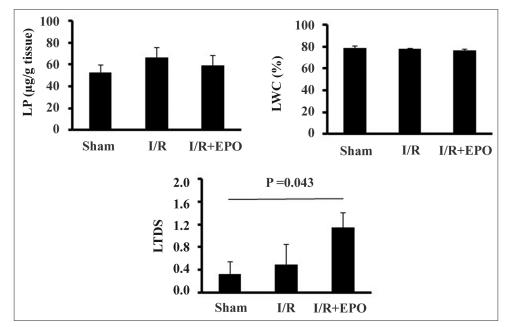
MDA is well-known as a final product of lipid peroxidation.<sup>[14]</sup> In our study, the serum and tissue levels of MDA increased non-significantly after renal IR. Previous studies have reported increased level of MDA after IRI.<sup>[14,45]</sup> However, in the study performed by Rasulian, no change in the serum level of MDA was reported; probably due to increased activity of super oxidase dismutase.<sup>[46]</sup>

The vasodilatory action of NO on vascular smooth muscle cells is well-known.<sup>[47]</sup> Decreased NO serum and tissue levels in the IR group may be related to reduced endothelial nitric oxide synthase (eNOS) in ischemic AKI.<sup>[48]</sup> On the other hand, EPO activates eNOS<sup>[13,47,49]</sup> that is agreement with our results for the EPO treated group. In the

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**Figure 2:** Kidney (k) and lung (l) tissue images (magnification ×100). K1-K3 and L1-L3 demonstrate the kidney and lung tissues image of groups 1-3. More tissue damages were observed in group 2 (K2 and L2). Erythropoietin indicates less kidney tissue damage (K3). However, it may promote the lung tissue damage (L3)



**Figure 3:** Lung endothelial permeability, lung water content, and lung tissue damage score in the sham, ischemia/reperfusion (I/R), and I/R treated with erythropoietin groups

current study, no change in LWC and LP were observed within 72 h after ischemia. Kramer *et al.* demonstrated that lung interstitial edema, vascular permeability occur 24 h and 48 h after renal IRI, but not 96 h after renal IRI. This is correlated with the level of Cr itself,<sup>[50]</sup> as we observed in the current study. In other study, pulmonary edema was observed after renal ischemia.<sup>[51]</sup> A limitation of

the current study is not measuring the parameters at day 2 after ischemia.

Lung tissue damage score (LTDS) was characterized by the presence of congestion, inflammatory cells, and hemorrhage; and EPO did not decrease its level. Wu *et al.* reported that low dose of EPO (300 IU/kg) increased lung injury after endotoxin shock via enhancing production of pro-inflammatory cytokines.<sup>[38,52]</sup> Furthermore, other study reported that low dose of EPO increase production of tumor necrosis factor- $\alpha$  and IL-6 in partial hepatectomy model.<sup>[53]</sup> These findings showed that EPO (500 IU/kg) probably induces lung tissue damage by activating pro-inflammatory pathways.

# CONCLUSIONS

The present study indicated that EPO have protective effects against renal injury induced by renal ischemia-reperfusion, but it did not improve the lung tissue damage induced by I/R injury.

# **ACKNOWLEDGMENTS**

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# REFERENCES

- 1. Bonventre JV, Yang L. Cellular pathophysiology of ischemic acute kidney injury. J Clin Invest 2011;121:4210-21.
- 2. Munshi R, Hsu C, Himmelfarb J. Advances in understanding ischemic acute kidney injury. BMC Med 2011;9:11.
- 3. Sharfuddin AA, Molitoris BA. Pathophysiology of ischemic acute kidney injury. Nat Rev Nephrol 2011;7:189-200.
- 4. Sheridan AM, Bonventre JV. Cell biology and molecular mechanisms of injury in ischemic acute renal failure. Curr Opin Nephrol Hypertens 2000;9:427-34.
- 5. Kribben A, Edelstein CL, Schrier RW. Pathophysiology of acute renal failure. J Nephrol 1999;12:S142-51.
- Sølling C, Christensen AT, Krag S, Frøkiaer J, Wogensen L, Krog J, *et al.* Erythropoietin administration is associated with short-term improvement in glomerular filtration rate after ischemia-reperfusion injury. Acta Anaesthesiol Scand 2011;55:185-95.
- Wang HB, Wang LM, Zhang RX, Liang R. Effects of erythropoietin on the expression of aquaporin-2 after renal ischemia-reperfusion injury: Experiment with rats. Zhonghua Yi Xue Za Zhi 2008;88:2710-4.

- Yazihan N, Kavas GO. Protective effect of erythropoietin in renal ischemia-reperfusion injury. Open Drug Discov J 2010;2:3-7.
- 9. Li X, Hassoun HT, Santora R, Rabb H. Organ crosstalk: The role of the kidney. Curr Opin Crit Care 2009;15:481-7.
- 10. Paladino JD, Hotchkiss JR, Rabb H. Acute kidney injury and lung dysfunction: A paradigm for remote organ effects of kidney disease? Microvasc Res 2009;77:8-12.
- 11. Ko GJ, Rabb H, Hassoun HT. Kidney-lung crosstalk in the critically ill patient. Blood Purif 2009;28:75-83.
- Rabb H, Wang Z, Nemoto T, Hotchkiss J, Yokota N, Soleimani M. Acute renal failure leads to dysregulation of lung salt and water channels. Kidney Int 2003;63:600-6.
- 13. Moore E, Bellomo R. Erythropoietin (EPO) in acute kidney injury. Ann Intensive Care 2011;1:3.
- 14. Kiris I, Kapan S, Kilbas A, Yilmaz N, Altuntaş I, Karahan N, *et al.* The protective effect of erythropoietin on renal injury induced by abdominal aortic-ischemia-reperfusion in rats. J Surg Res 2008;149:206-13.
- 15. Ates E, Yalcin AU, Yilmaz S, Koken T, Tokyol C. Protective effect of erythropoietin on renal ischemia and reperfusion injury. ANZ J Surg 2005;75:1100-5.
- 16. Pallet N, Rabant M, Legendre C, Martinez F, Choukroun G. The nephroprotective properties of recombinant human erythropoietin in kidney transplantation: Experimental facts and clinical proofs. Am J Transplant 2012;12:3184-90.
- 17. Ardalan MR, Estakhri R, Hajipour B, Ansarin K, Asl NA, Nasirizade MR, *et al.* Erythropoietin ameliorates oxidative stress and tissue injury following renal ischemia/reperfusion in rat kidney and lung. Med Princ Pract 2013;22:70-4.
- Cassis P, Gallon L, Benigni A, Mister M, Pezzotta A, Solini S, *et al.* Erythropoietin, but not the correction of anemia alone, protects from chronic kidney allograft injury. Kidney Int 2012;81:903-18.
- 19. Kotecha R, Toledo-Pereyra LH. The renal protective effect of erythropoietin on acute ischemic injury in kidney transplantation. J Surg Res 2012;178:611-3.
- 20. Song YR, Lee T, You SJ, Chin HJ, Chae DW, Lim C, *et al.* Prevention of acute kidney injury by erythropoietin in patients undergoing coronary artery bypass grafting: A pilot study. Am J Nephrol 2009;30:253-60.
- Sureshkumar KK, Hussain SM, Ko TY, Thai NL, Marcus RJ. Effect of high-dose erythropoietin on graft function after kidney transplantation: A randomized, double-blind clinical trial. Clin J Am Soc Nephrol 2012;7:1498-506.
- 22. Oh SW, Chin HJ, Chae DW, Na KY. Erythropoietin improves long-term outcomes in patients with acute kidney injury after coronary artery bypass grafting. J Korean Med Sci 2012;27:506-11.

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- 23. Aydin Z, Mallat MJ, Schaapherder AF, van Zonneveld AJ, van Kooten C, Rabelink TJ, *et al.* Randomized trial of short-course high-dose erythropoietin in donation after cardiac death kidney transplant recipients. Am J Transplant 2012;12:1793-800.
- 24. Souza AC, Volpini RA, Shimizu MH, Sanches TR, Camara NO, Semedo P, *et al*. Erythropoietin prevents sepsis-related acute kidney injury in rats by inhibiting NF-κB and upregulating endothelial nitric oxide synthase. Am J Physiol Renal Physiol 2012;302:F1045-54.
- 25. Yang FL, Subeq YM, Chiu YH, Lee RP, Lee CJ, Hsu BG. Recombinant human erythropoietin reduces rhabdomyolysis-induced acute renal failure in rats. Injury 2012;43:367-73.
- 26. Caetano AM, Vianna Filho PT, Castiglia YM, Golim MA, de Souza AV, de Carvalho LR, *et al.* Erythropoietin attenuates apoptosis after ischemia-reperfusion-induced renal injury in transiently hyperglycemic Wister rats. Transplant Proc 2011;43:3618-21.
- 27. Hu L, Yang C, Zhao T, Xu M, Tang Q, Yang B, *et al.* Erythropoietin ameliorates renal ischemia and reperfusion injury via inhibiting tubulointerstitial inflammation. J Surg Res 2012;176:260-6.
- 28. Cassis P, Azzollini N, Solini S, Mister M, Aiello S, Cugini D, *et al.* Both darbepoetin alfa and carbamylated erythropoietin prevent kidney graft dysfunction due to ischemia/ reperfusion in rats. Transplantation 2011;92:271-9.
- 29. Prókai A, Fekete A, Bánki NF, Müller V, Vér A, Degrell P, *et al.* Renoprotective effect of erythropoietin in rats subjected to ischemia/reperfusion injury: Gender differences. Surgery 2011;150:39-47.
- 30. Ishii Y, Sawada T, Murakami T, Sakuraoka Y, Shiraki T, Shimizu A, *et al*. Renoprotective effect of erythropoietin against ischaemia-reperfusion injury in a non-human primate model. Nephrol Dial Transplant 2011;26:1157-62.
- 31. Moriyama MT, Tanaka T, Morita N, Ishii T, Chikazawa I, Suga K, *et al.* Renal protective effects of erythropoietin on ischemic reperfusion injury. Cell Transplant 2010;19:713-21.
- 32. Bahlmann FH, Fliser D. Erythropoietin and renoprotection. Curr Opin Nephrol Hypertens 2009;18:15-20.
- 33. Wu H, Ren B, Zhu J, Dong G, Xu B, Wang C, *et al.* Pretreatment with recombined human erythropoietin attenuates ischemia-reperfusion-induced lung injury in rats. Eur J Cardiothorac Surg 2006;29:902-7.
- Mänttäri M, Mälkönen M, Manninen V. Effect of diazepam on endothelial permeability, plasma lipids and lipoproteins in cholesterol fed rabbits. Acta Med Scand Suppl 1982;660:109-13.
- 35. Nematbakhsh M, Hayat-Davoodi P, Rajabi P, Samarian SH. The effect of estrogen on endothelial permeability of aorta and the level of serum nitrite

concentration in cholesterol-fed ovariectomized rabbit. Iran Biomed J 2002;6:77-82.

- 36. Spandou E, Tsouchnikas I, Karkavelas G, Dounousi E, Simeonidou C, Guiba-Tziampiri O, *et al.* Erythropoietin attenuates renal injury in experimental acute renal failure ischaemic/reperfusion model. Nephrol Dial Transplant 2006;21:330-6.
- 37. Sharples EJ, Patel N, Brown P, Stewart K, Mota-Philipe H, Sheaff M, *et al*. Erythropoietin protects the kidney against the injury and dysfunction caused by ischemia-reperfusion. J Am Soc Nephrol 2004;15:2115-24.
- Wu WT, Hu TM, Lin NT, Subeq YM, Lee RP, Hsu BG. Low-dose erythropoietin aggravates endotoxin-induced organ damage in conscious rats. Cytokine 2010;49:155-62.
- Forbes JM, Hewitson TD, Becker GJ, Jones CL. Ischemic acute renal failure: Long-term histology of cell and matrix changes in the rat. Kidney Int 2000;57:2375-85.
- 40. Johnson DW, Pat B, Vesey DA, Guan Z, Endre Z, Gobe GC. Delayed administration of darbepoetin or erythropoietin protects against ischemic acute renal injury and failure. Kidney Int 2006;69:1806-13.
- 41. Patel NS, Sharples EJ, Cuzzocrea S, Chatterjee PK, Britti D, Yaqoob MM, *et al.* Pretreatment with EPO reduces the injury and dysfunction caused by ischemia/ reperfusion in the mouse kidney *in vivo*. Kidney Int 2004;66:983-9.
- 42. Vesey DA, Cheung C, Pat B, Endre Z, Gobé G, Johnson DW. Erythropoietin protects against ischaemic acute renal injury. Nephrol Dial Transplant 2004;19:348-55.
- 43. Hassoun HT, Grigoryev DN, Lie ML, Liu M, Cheadle C, Tuder RM, *et al.* Ischemic acute kidney injury induces a distant organ functional and genomic response distinguishable from bilateral nephrectomy. Am J Physiol Renal Physiol 2007;293:F30-40.
- 44. Nemoto T, Yokota N, Keane WF, Rabb H. Recombinant erythropoietin rapidly treats anemia in ischemic acute renal failure. Kidney Int 2001;59:246-51.
- 45. Sener G, Sehirli AO, Keyer-Uysal M, Arbak S, Ersoy Y, Yeğen BC. The protective effect of melatonin on renal ischemia-reperfusion injury in the rat. J Pineal Res 2002;32:120-6.
- 46. Rasoulian B, Jafari M, Noroozzadeh A, Mehrani H, Wahhab-Aghai H, Hashemi-Madani S, *et al.* Effects of ischemia-reperfusion on rat renal tissue antioxidant systems and lipid peroxidation. Acta Med Iran 2008;46:353-60.
- 47. Rezaeian F, Wettstein R, Egger JF, Sandmann F, Rücker M, Tobalem M, *et al.* Erythropoietin-induced upregulation of endothelial nitric oxide synthase but not vascular endothelial growth factor prevents musculocutaneous tissue from ischemic damage. Lab Invest 2010;90:40-51.

- Betz B, Schneider R, Kress T, Schick MA, Wanner C, Sauvant C. Rosiglitazone affects nitric oxide synthases and improves renal outcome in a rat model of severe ischemia/ reperfusion injury. PPAR Res 2012;2012:219319.
- 49. Oba S, Suzuki E, Nishimatsu H, Kumano S, Hosoda C, Homma Y, *et al.* Renoprotective effect of erythropoietin in ischemia/reperfusion injury: Possible roles of the Akt/ endothelial nitric oxide synthase-dependent pathway. Int J Urol 2012;19:248-55.
- Kramer AA, Postler G, Salhab KF, Mendez C, Carey LC, Rabb H. Renal ischemia/reperfusion leads to macrophage-mediated increase in pulmonary vascular permeability. Kidney Int 1999;55:2362-7.
- 51. Deng J, Hu X, Yuen PS, Star RA. Alpha-melanocyte-

stimulating hormone inhibits lung injury after renal ischemia/reperfusion. Am J Respir Crit Care Med 2004;169:749-56.

- 52. Kakavas S, Demestiha T, Vasileiou P, Xanthos T. Erythropoetin as a novel agent with pleiotropic effects against acute lung injury. Eur J Clin Pharmacol 2011;67:1-9.
- 53. Klemm K, Eipel C, Cantré D, Abshagen K, Menger MD, Vollmar B. Multiple doses of erythropoietin impair liver regeneration by increasing TNF-alpha, the Bax to Bcl-xL ratio and apoptotic cell death. PLoS One 2008;3:e3924.

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